



Book of Abstracts. The 1st Next Generation Conversations: Albany at Ruston 2024 (The 21st Albany Conversation)

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Book of Abstracts. The 1st Next Generation Conversations: Albany at Ruston 2024 (The 21st Albany Conversation)

Prepared and edited by

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Conversation starters

The original Conversation hosted by Prof. Ramaswamy Sarma was published as *Stereodynamics of Molecular Systems: Proceedings of a symposium held at the State University of New York at Albany 23–24 April 1979*. Sarma undertook the task of inviting scientists and students alike from all over the world to a remote college campus to discuss science. Against all odds, the idea caught on, leading to a conference series that enjoyed a run of 20 meetings spanning four decades with several hundred participants at its zenith. Students and Nobel Laureates alike partook of lively talks and expansive poster sessions, enjoying the comradery of the Conversations in a setting devoid of distractions. Well ahead of its time, the conference boasted a diversity

that cut across race, class, gender, academic rank, and country of origin, no small feat in the climate of the Cold War Era; the conference drew a notable following of Indian, Russian, Israeli, and Western European participants to this remote location all unified by one simple thing: they desired to engage in open dialogue about the full S.E.T. (simulation, experiment, theory) of approaches to biomolecular science.

After running twenty celebrated Conversations, Prof. Sarma appointed Thomas C. Bishop at Louisiana Tech University in Ruston, LA to continue the series. On 11–15 June 2024, a new vision of the Next Gen Conversations was realized with Kelly M. Thayer of Wesleyan University, Middletown, CT and Robert T. Young, Rutgers University, Piscataway, NJ joining the



Figure 1. Attendees of the 1st NextGen Conversations: Albany @ Ruston. Photo credit: Estevan Garcia, Louisiana Tech University, Ruston, LA.

organizing committee (Figure 1). Participants hailed from two continents at the inaugural event. Six sessions were held covering various topics of biomolecular science across three days. Posters showcased additional student work. The week culminated with the keynote address delivered by Prof. Wilma K. Olson, Rutgers University, Piscataway, NJ, a leader in the field of nucleic acids who also attended every Albany Conversation. She entwined the history of DNA structure, snapshots from previous conferences, and fresh research results, through adenine, one of the four nucleotides. When seasoned Conversation attenders reconvened, the murmur quickly grew into the same lively question and answer sessions that typified the Albany experience. The students were also actively involved in the Conversation: showing posters, discussing science and career advancement over lunch, dinner, or the picnic table in front of the 'Hospitality Suite' at the dorms. New faces, new surroundings, and new organizers ushered in the Next Generation of the Conversations, but not without the essence of what made an entirely unlikely concept of geographically remote meetings of world class scientists such a success.

This collection of Abstracts not only attests to the science comprising the conference, but also heralds the re-envisioning of the Conversations set squarely upon the monumental foundation of the prior twenty Conversations. On behalf of the thousands of scientists that attended the four decades of Conversations, we express our deep appreciation to Prof. Sarma for growing a seemingly impossible vision into a conference series that profoundly impacted innumerable research projects, careers, and participants scattered across the globe. This collection celebrates the rekindling of the Conversations anew. The organizers extend an invitation to join the 2nd Next Gen Conversations, June 2026 in Ruston, LA.

The organizers gratefully acknowledge Prof. Aditya Mittal for his hand in perpetuating the Conversations, garnering enthusiasm among the Indian participants, and scientific contributions. We thank Louisiana Tech University for hosting the event, and their numerous staff and student workers who made this event possible.

This Book of Abstracts is dedicated to Prof. Ramaswamy Sarma for initiating and perpetuating an influential internationally acclaimed conference series on biomolecular science for forty years.




Abstracts

1. An adenine story (2024 keynote address)

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KEYWORDS

Adenine; DNA sequence-dependent structure; noncanonical RNA base pairs

The story begins¹ with my own introduction to nucleic acids through polyriboadenylic acid and the development of a molecular model that takes account of its temperature-dependent biophysical properties (Eisenberg & Felsenfeld, 1967; Olson & Flory, 1972). The discussion then turns to early ideas on the structural mechanisms responsible for the curvature of double-helical DNA bearing repeated tracts of A's (Marini et al., 1982), such as the local kinks hypothesized to occur in nucleosomal DNA (Crick & Klug, 1975; Trifonov & Sussman, 1980) and the observed abutment of different helical forms in DNA block copolymers (Early et al., 1977). The large number of high-resolution structures now at hand makes clear the contributions of nucleotide sequence to DNA structure, including the preferred configurations of the ten unique base pair steps and the influence of their immediate neighbors on local structure (Olson et al., 2024). The methodology used to characterize the arrangements of successive base pairs can also be applied to study the edge-to-edge associations of non-canonical base pairs and the structural context in which they occur (Lu et al., 2015), such as the tertiary A-minor motifs that bring adenines in direct contact with the minor-groove edges of canonical base pairs in distant parts of RNA structures (Nissen et al., 2001). The story concludes with a discussion of the long stretch of A•A base pairs found in the *Escherichia coli* ribosome and the unusual folded architecture in which they occur.

Acknowledgments

I am grateful to the students and colleagues who contributed over the years to this work.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

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¹The following are 28 abstracts presented in alphabetical order according to the first author's last name. Two of the authors were unable to attend the meeting in person; we nonetheless present their abstracts *in absentia*. For each abstract, the presenter's name is underlined, and their email address provided.

References

- Crick, F. H., & Klug, A. (1975). Kinky helix. *Nature*, 255(5509), 530–533. doi: [10.1038/255530a0](https://doi.org/10.1038/255530a0)
- Early, T. A., Kearns, D. R., Burd, J. F., Larson, J. E., & Wells, R. D. (1977). High resolution proton nuclear magnetic resonance investigation of the structural and dynamic properties of d(C15A15)-d(T15G15). *Biochemistry*, 16(3), 541–551. doi: [10.1021/bi00622a031](https://doi.org/10.1021/bi00622a031)
- Eisenberg, H., & Felsenfeld, G. (1967). Studies of the temperature-dependent conformation and phase separation of polyriboadenylic acid solutions at neutral pH. *Journal of Molecular Biology*, 30(1), 17–37. doi: [10.1016/0022-2836\(67\)90240-9](https://doi.org/10.1016/0022-2836(67)90240-9)
- Lu, X.-J., Bussemaker, H. J., & Olson, W. K. (2015). DSSR: An integrated software tool for dissecting the spatial structure of RNA. *Nucleic Acids Research*, 43(21), e142. doi: [10.1093/nar/gkv716](https://doi.org/10.1093/nar/gkv716)
- Marini, J. C., Levene, S. D., Crothers, D. M., & Englund, P. T. (1982). Bent helical structure in kinetoplast DNA. *Proceedings of the National Academy of Sciences of the United States of America*, 79(24), 7664–7668. doi: [10.1073/pnas.79.24.7664](https://doi.org/10.1073/pnas.79.24.7664)
- Nissen, P., Ippolito, J. A., Ban, N., Moore, P. B., & Steitz, T. A. (2001). RNA tertiary interactions in the large ribosomal subunit: The A-minor motif. *Proceedings of the National Academy of Sciences of the United States of America*, 98(9), 4899–4903. doi: [10.1073/pnas.081082398](https://doi.org/10.1073/pnas.081082398)
- Olson, W. K., & Flory, P. J. (1972). Spatial configuration of polynucleotide chains. II. Conformational energies and the average dimensions of polyribo-nucleotides. *Biopolymers*, 11(1), 25–56. doi: [10.1002/bip.1972.360110103](https://doi.org/10.1002/bip.1972.360110103)
- Olson, W. K., Young, R. T., & Czapla, L. (2024). DNA simulation benchmarks revealed with the accumulation of high-resolution structures. *Biophysical Reviews*, 16, 275–284. doi: [10.1007/s12551-024-01198-2](https://doi.org/10.1007/s12551-024-01198-2)
- Trifonov, E. N., & Sussman, J. L. (1980). The pitch of chromatin DNA is reflected in its nucleotide sequence. *Proceedings of the National Academy of Sciences of the United States of America*, 77(7), 3816–3820. doi: [10.1073/pnas.77.7.3816](https://doi.org/10.1073/pnas.77.7.3816)

2. [*In absentia*] Correlation function for heteropolymers

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KEYWORDS

Heteropolymers; biopolymers; correlation function

The correlation function between helical states in two-component (A/B) random biopolymers has been investigated within the framework of the Generalized Model of Polypeptide Chain (GMPC). Correlation function 'g' was computed as a function of distance 'D' separating helical states. The transfer matrix of GMPC is used as in. Partition function 'Z' and the helicity degree 'θ' are calculated. Correlation function results are obtained for the melting temperature, and boundaries within the melting interval (see Figure 1). Based on dependencies of $\ln(g(D))$ on D and $\ln(g(D))$ on $\ln(D)$, we propose an analytical relationship for 'g' and 'D': $g(D) = Ae^{-D/\xi} + BD - n$. Here 'A', 'B' and 'n', 'ξ' are constants estimated through logarithmic graphs, and the first term on right hand side corresponds to homopolymer dependence with correlation length ξ. The second term has been used for spin-glass models. The $g(D)$ curve in melting temperature

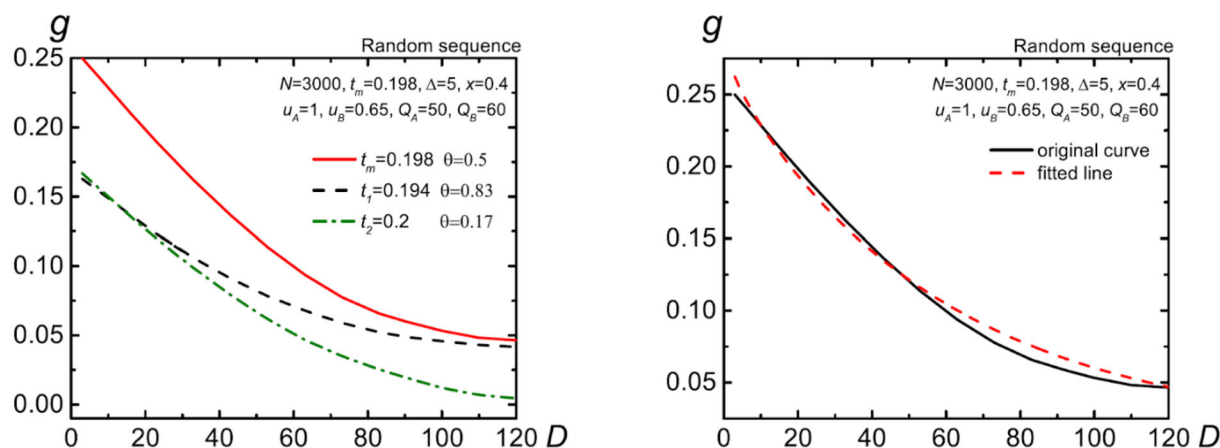


Figure 1. $g(D)$ dependence in melting temperature: $t(\theta = 0.5) = 0.198$, and in boundary temperatures of melting interval: $t_1(\theta = 0.17) = 0.2$, $t_2(\theta = 0.83) = 0.194$ (left), fitting curve for melting temperature (right), with constants: $A \approx 0.24$, $B \approx 0.04$, $\zeta \approx 59$, $n = 0.21$.

was fitted with constants using the formula: $g(D)$. For additional information see Asatryan et al., 2022; Flory, 1969; Gunnarsson et al., 1988; Tonoyan et al., 2015.

Conclusions

- The exponential dependence of the correlation function predominates near the melting temperature, which explains the applicability of homopolymer models for DNA.
- The small magnitude of the maximum correlation length ξ in contrast to the values for the corresponding homopolymers is consistent with the broadening of the transition interval for heteropolymers.
- The presence of the order component in dependence for long tails indicates correlations associated with the heterogeneity of the system and needs further research.

Disclosure statement

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References

- Asatryan, A. V., Mikayelyan, H. H., & Stepanyan, V. A. (2022). Helix-coil transition in heterogeneous biopolymers: Influence of fixing bond scale. *Journal of Contemporary Physics*, 57(3), 308–312. doi: [10.1134/S1068337222030057](https://doi.org/10.1134/S1068337222030057)
- Flory, P. J. (1969). *Statistical mechanics of chain molecules*. Interscience. doi: [10.1002/bip.1969.360080514](https://doi.org/10.1002/bip.1969.360080514)
- Gunnarsson, K., Svedlindh, P., Nordblad, P., Lundgren, L., Aruga, H., & Ito, A. (1988). Dynamics of an Ising spin-glass in the vicinity of the spin-glass temperature. *Physical Review Letters*, 61(6), 754–757. doi: [10.1103/PhysRevLett.61.754](https://doi.org/10.1103/PhysRevLett.61.754)
- Tonoyan, S., Asatryan, A. V., Andriasyan, A. K., Mamasakhlisov, Y., & Morozov, V. F. (2015). Helix-coil transitions in heteropolymers: The constrained annealing approach. *Journal of Biomolecular Structure & Dynamics*, 33(sup1), 126–126. doi: [10.1080/07391102.2015.1032828](https://doi.org/10.1080/07391102.2015.1032828)

3. Reconstruction of a dynamic process: stages in non-enveloped virus disassembly

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KEYWORDS

Non-enveloped virus; disassembly; thermostability; cryo-electron microscopy

Non-enveloped icosahedral viruses have a stable and symmetric protein shell, or capsid, which encapsulates the genome (Figure 1). While stability of the shell is crucial for protecting and transporting the viral genome; dynamic behavior of specific capsid components is also required during cellular membrane penetration and viral genome release in the early stages of entry (Kumar et al., 2018). We are attempting to establish the molecular pathways of disassembly for non-enveloped viruses using a model system—a $T=3$ icosahedral insect virus, Flock House Virus (FHV) (Bajaj et al., 2016). *In vitro* heat-induced disassembly of infectious FHV particles indicated stepwise alterations in capsid conformation leading to disassembly, which is not evident in non-infectious mutated versions of the same particle (Azad & Banerjee, 2019). Cryo-electron microscopy and single particle reconstructions of disassembling particles indicated conformational alterations including ‘puffing’ of particles triggered by movement of subunit proteins, and major alterations at symmetry axes. Asymmetric reconstructions indicated directional genome release from particles, which suggests structural differences in sequentially identical capsid proteins occupying different positions in the icosahedral asymmetric units of the capsid (Azad & Banerjee, 2019). All atom simulations of the whole capsid supported the existence of favored pathways within the capsid for externalization of flexible components and the genome (Shrivastav et al., 2024). A combination of

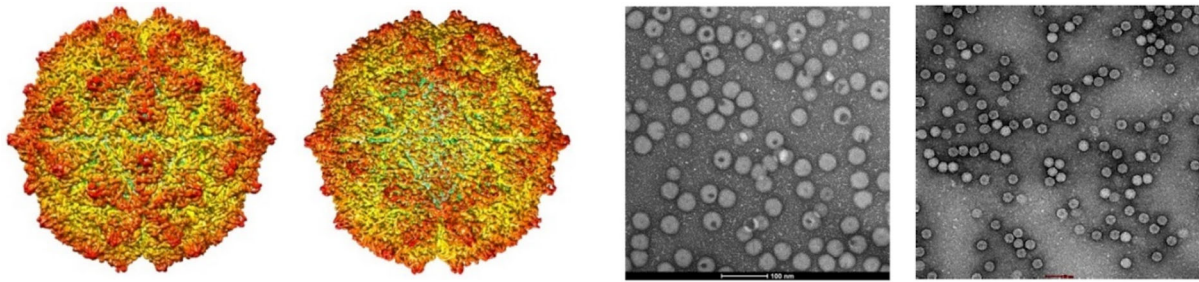


Figure 1. CryoEM single particle reconstructions and negative stain images of FHV disassembly intermediates.

heat induced disassembly and cryoEM was utilized to study the disassembly of two other non-enveloped icosahedral viruses—a mammalian RNA virus—Feline Calicivirus (FCV), and a double-stranded DNA virus—Human Adenovirus 5 (HAdV5). Although these viruses are fairly different in genome content and size from FHV, they appear to undergo similar stepwise disassembly-related conformational alterations, with striking morphological similarities between at least one disassembly intermediate of FHV and FCV. It is hoped that cryoEM and simulation studies can be combined to create a molecular roadmap for the stages in disassembly. Since non-enveloped viruses typically display higher resistance to disinfectants and sanitizers compared to their enveloped counterparts; a molecular level understanding of disassembly pathways is required for devising globally effective chemical inactivation strategies.

Acknowledgments

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Disclosure statement

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References

- Azad, K., & Banerjee, M. (2019). Structural dynamics of nonenveloped virus disassembly intermediates. *Journal of Virology*, *93*(22), 1115–1119. doi: [10.1128/JVI.01115-19](https://doi.org/10.1128/JVI.01115-19)
- Bajaj, S., Dey, D., Bhukar, R., Kumar, M., & Banerjee, M. (2016). Non-enveloped virus entry: structural determinants and mechanism of functioning of a viral lytic peptide. *Journal of Molecular Biology*, *428*(17), 3540–3556. doi: [10.1016/j.jmb.2016.06.006](https://doi.org/10.1016/j.jmb.2016.06.006)
- Kumar, C. S., Dey, D., Ghosh, S., & Banerjee, M. (2018). Breach: Host membrane penetration and entry by nonenveloped viruses. *Trends in Microbiology*, *26*(6), 525–537. doi: [10.1016/j.tim.2017.09.010](https://doi.org/10.1016/j.tim.2017.09.010)
- Shrivastav, G., Borkotoky, S., Dey, D., Singh, B., Malhotra, N., Azad, K., Jayaram, B., Agarwal, M., & Banerjee, M. (2024). Structure and energetics guide dynamic behaviour in a $T=3$ icosahedral virus capsid. *Biophysical Chemistry*, *305*, 107152. doi: [10.1016/j.bpc.2023.107152](https://doi.org/10.1016/j.bpc.2023.107152)

4. Recycling the sequence, structure, function paradigm for chromatin

Thomas Connor Bishop

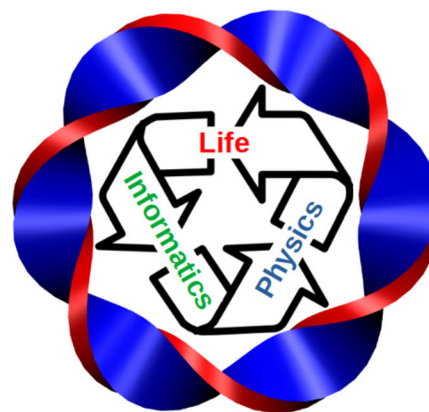
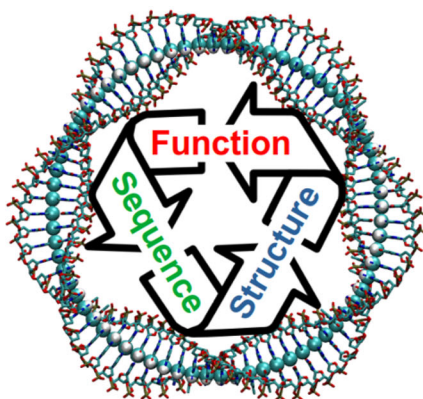
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KEYWORDS

Chromatin; space curves; genome dashboard; DNA information technology

The sequence-structure-function paradigm is an established model for assessing protein folding that has been recycled



for chromatin folding. Superficially this is merely updating an old idea. Here, I argue that unifying the sequence-structure-function paradigm is a necessary step in advancing our understanding of chromatin. Historically bioinformatics/sequence analysis and structural/computational biology are distinct scientific disciplines with different experimental methods, scientific theories, and computational requirements. Bioinformatics is based on information theory, and sequence is considered a mathematical concept regarding the one-dimensional ordering of elements in a set. Structural biology is based on the laws of physics and is concerned with measuring or modeling the spatial arrangement of atoms, residues, and molecules in three-dimensional space. The purpose of our genome dashboards is to unify these approaches and fully integrate sequence and structure data (Li et al., 2020). This alone provides a novel means for validating data and generating hypotheses. The mathematical transform between 1D and 3D representations is achieved by consideration of geometrically exact space curves and generalization of the DNA base pair step parameters. The transform is inherently multi-scale and may provide a basis for the development of Richardson Diagrams for chromatin. The function of chromatin is to support life processes, but within the genome dashboard framework function is extensible. Chromatin may encode books, movies, software, and more.

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Reference

Li, Z., Sun, R., & Bishop, T. C. (2020). Genome dashboards: Framework and examples. *Biophysical Journal*, 118(9), 2077–2085. doi: [10.1016/j.bpj.2020.02.018](https://doi.org/10.1016/j.bpj.2020.02.018)

5. Tumor suppressor protein p53 isoform dynamics

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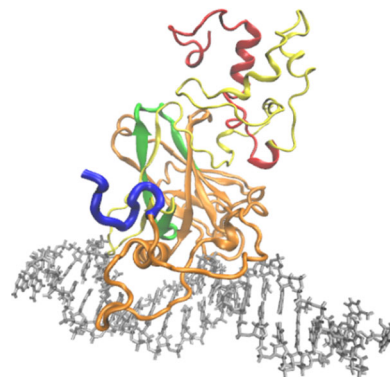
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KEYWORDS

DNA; p53; splice variants; mutations

Many eukaryotic proteins are translated from their encoding DNA not only as a single sequence but instead as several variants arising from the processing of splice variants. The



tumor suppressor protein p53, a key drug target for curing cancer, exhibits a dozen naturally occurring splice variants, yet the majority of current studies do not take this into account. To bridge this gap, we explore the hypothesis that the difference in sequence may hard code differences in p53-DNA interactions, affecting its role as a transcription factor by considering a variety of naturally occurring sequence variants of this protein. We postulate that the sequence variations may inherently regulate p53 activity, and we seek to characterize these dynamic differences, which may prove crucial for developing effective cancer treatments (Armour-Garb et al., 2022). We present a comparison of these properties using molecular dynamics simulations. Taking this a step further, we explore the effects of mutations on the isoforms. In particular, we investigate Y220C mutation on various isoforms of the p53 protein from a dynamic standpoint.

Disclosure statement

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Reference

Armour-Garb, I., Han, I. S. M., Cowan, B. S., & Thayer, K. M. (2022). Variable regions of p53 isoforms allosterically hard code DNA interaction. *The Journal of Physical Chemistry. B*, 126(42), 8495–8507. doi: [10.1021/acs.jpccb.2c06229](https://doi.org/10.1021/acs.jpccb.2c06229).

6. Tracing the birth and intrinsic disorder of loops and domains in protein evolution

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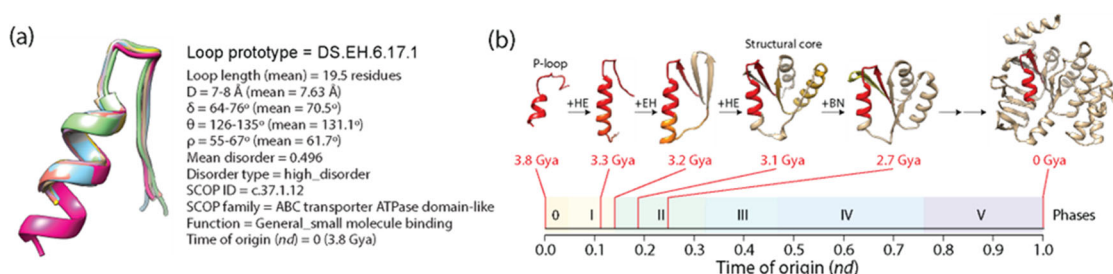


Figure 1. The P-loop, the most ancient loop prototype (a), and the birth of the ATP-binding ABC transporter domain family (b). Gya: billions of years ago sensu a clock of folds; nd: node distance in phylogenomic tree.

KEYWORDS

Constraints; evolution; intrinsic disorder; protein loops; structural domains

Protein loops and structural domains are molecular building blocks—prior molecular states. They are largely responsible for the processes and molecular functions of the living world. Here, we focus on the emergence and evolution of these elemental architects of protein structure and the primordial role of intrinsic disorder. Phylogenomic reconstruction spanning super kingdoms and viruses generated evolutionary chronologies of loop prototypes sourced from ArchDB and SCOP domains defining six distinct evolutionary phases and a most parsimonious evolutionary progression of cellular life. Each phase was marked by strategic loop and domain accumulation shaping the structures and functions of common ancestors. Chronologies allowed us to trace the birth of domain structures by recruitment of loops, which we modeled *ab initio* with AlphaFold2 (Figure 1). A phylogenomic survey of disorder in loops and domains revealed that loop-associated ‘short’ and domain-associated ‘long’ regions of disorder evolved differently across different levels of protein organization. Ancient loops tended to be more disordered than their derived counterparts, highlighting the central evolutionary role of disorder and flexibility. In contrast, ancient domains were ordered, with disorder evolving as a benefit acquired later in evolution. Percolation of evolutionary constraints from higher to lower levels of organization resulted in trade-offs between flexibility and rigidity impacting loop structure and geometry. An exploration of loop geometric properties revealed gradual replacement of prototypes with α -helix and β -strand bracing structures over time, paving the way for the dominance of other loop types. Our findings provide a deep evolutionary view of the link between structure, disorder, flexibility, and function, as well as insights into the evolutionary role of intrinsic disorder and its contribution to protein structure and function. For additional information see Aziz et al., 2016, 2023; Mughal & Caetano-Anollés, 2023.

Disclosure statement

No potential conflict of interest was reported by the author(s).

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References

- Aziz, M. F., Caetano-Anollés, K., & Caetano-Anollés, G. (2016). The early history and emergence of molecular functions and modular scale-free network behavior. *Scientific Reports*, 6(1), 25058. doi: [10.1038/srep25058](https://doi.org/10.1038/srep25058)
- Aziz, M. F., Mughal, F., & Caetano-Anollés, G. (2023). Tracing the birth of structural domains from loops during protein evolution. *Scientific Reports*, 13(1), 14688. doi: [10.1038/s41598-023-41556-w](https://doi.org/10.1038/s41598-023-41556-w).
- Caetano-Anollés, K., Aziz, M. F., Mughal, F., & Caetano-Anollés, G. (2024). On protein loops, prior molecular states and common ancestors of life. *Journal of Molecular Evolution*. doi: [10.1007/s00239-024-10167-y](https://doi.org/10.1007/s00239-024-10167-y).
- Mughal, F., & Caetano-Anollés, G. (2023). Evolution of intrinsic disorder in protein loops. *Life*, 13(10), 2055. doi: [10.3390/life13102055](https://doi.org/10.3390/life13102055).

7. Synthesis of oxaliplatin analogs and their controlled release from hydrogels

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KEYWORDS

Oxaliplatin; hydrogel; drug delivery; simvastatin

Platinum based chemotherapeutics are widely used to treat multiple types of cancer (Figure 1). To improve chemotherapy, it is important to reduce their harmful side effects while maintaining and potentially increasing their efficacy. We have previously developed a photo-crosslinked hydrogel for local drug delivery and have achieved sustained release of simvastatin (SMV), a hydrophobic statin drug. Recently, statin drugs have been noted to enhance the antitumor effect against various types of cancer cells. In this work, we synthesized oxaliplatin analogs, performed cytotoxicity studies to test their potency, and incorporated them along with SMV into hydrogel systems using various methods, such as passive loading, micelle encapsulation, and platinum-ligand coordination binding. Alpha lipoic acid (ALA) is a naturally occurring antioxidant that has been reported to prevent cisplatin-induced adverse cytotoxicity. Several oxaliplatin analogs including ALA-oxaliplatin conjugate (OXA-ALA2) were synthesized and characterized *via* ¹H NMR, ¹⁹⁵Pt NMR, and mass spectrometry. Cytotoxicity tests

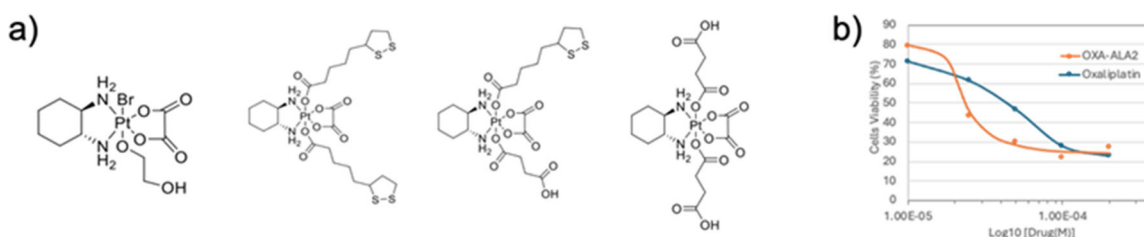


Figure 1. (a) Oxaliplatin analogs. (b) Cell viability results of OXA-ALA2 against MG-63.

were performed on MG-63 and SAOS-2 human osteosarcoma cells. OXA-ALA2 showed high cytotoxicity against these osteosarcoma cell lines, comparable to oxaliplatin. In addition, SMV exhibited strong cytotoxicity toward MG-63, indicating the potential to amplify the efficacy of oxaliplatin analogs in a dual drug delivery system. The antitumor effects of oxaliplatin will potentially be enhanced with the created analogs while simultaneously decreasing the adverse side effects by using injectable/implantable hydrogels in a controlled manner as an alternative delivery method. For additional information see Branca et al., 2021; Cisneros et al., 2021; Spengler et al., 1985.

Disclosure statement

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References

- Branca, J. J., Carrino, D., Gulisano, M., Ghelardini, C., Di Cesare Mannelli, L., & Pacini, A. (2021). Oxaliplatin-induced neuropathy: Genetic and epigenetic profile to better understand how to ameliorate this side effect. *Frontiers in Molecular Biosciences*, 8, 643824. doi: [10.3389/fmolb.2021.643824](https://doi.org/10.3389/fmolb.2021.643824).
- Cisneros, K., Chowdhury, N., Coleman, E., Ferdous, T., Su, H., Jennings, J. A., Bumgardner, J. D., & Fujiwara, T. (2021). Long-term controlled release of simvastatin from photoprinted triple-networked hydrogels composed of modified chitosan and PLA-Peg micelles. *Macromolecular Bioscience*, 27(8), e2100123. doi: [10.1002/mabi.202100123](https://doi.org/10.1002/mabi.202100123).

8. Landscapes of genomic architecture across evolution

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The human genome is composed of 46 DNA molecules—the chromosomes—with a combined length of about two meters. Chromosomes are stored in the cell nucleus in a very organized

fashion that is specific to the cell type and phase of life; this three-dimensional architecture is a key element of transcriptional regulation, and its disruption often leads to disease. What is the physical mechanism leading to genome architecture? If the DNA contained in every human cell is identical, where is the blueprint of such architecture stored. In this talk, I will demonstrate how the architecture of interphase chromosomes is encoded in the one-dimensional sequence of epigenetic markings much as three-dimensional protein structures are determined by their one-dimensional sequence of amino acids. In contrast to the situation for proteins, however, the sequence code provided by the epigenetic marks that decorate the chromatin fiber is not fixed but is dynamically rewritten during cell differentiation, modulating both the three-dimensional structure and gene expression in different cell types. This idea led to the development of a physical theory for the folding of genomes, which enables predicting the spatial conformation of chromosomes with unprecedented accuracy and specificity. Finally, I will demonstrate how the different physical processes in our model impact the topology of chromosomes across evolution.

9. [In absentia] Identification of iodothyronines in plant tissues

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KEYWORDS

Higher plants; iodothyronines; thyroxine; triiodothyronine; spectrophotometry; mass spectrometry; iodine content

Currently, there is no doubt that many of the signaling molecules are common to organisms of various systematic groups. This is probably true for such important metabolic regulators as iodothyronines. A number of studies have proven the presence of thyroid hormone activity in compounds of plant origin (see References). Nevertheless, based on the conducted studies, it is impossible to conclude whether the compounds under consideration, like animal and human thyroid hormones, are iodine derivatives of tyronine or whether they are mimetics of thyroid hormones. The purpose of this study was to find out whether iodine-thyronin analogues with different degrees of iodination are present in plant tissues, as well as to determine the concentration of iodine in plant tissue lysates and compare it with

theoretically calculated in accordance with the concentration of the studied compounds and the assumption of a structure identical to human thyroid hormones. It has been shown that potato tubers and wheat leaves simultaneously contain analogues of tetraiodothyronine (T4) and triiodothyronine (T3). In potato tubers at rest, the concentration of T4 was 118 ± 16 nmol/l ($n = 15$), in the same samples, the concentration of T3 was 4.01 ± 0.96 nmol/l. The concentrations of T4 and T3 in wheat leaf lysates were 60.24 ± 79 and 6.76 nmol/l ($n = 15$), respectively. By inductively coupled plasma mass spectrometry, it was found that the amount of iodine present in the studied samples corresponds to the assumption that the activity is due to the presence of tetraiodated derivatives of tyronine. For additional information see Bashkatov & Garipova, 2022; Bhattacharya et al., 2023; Chiellini et al., 2002; Garipova & Usmanova, 2013; Garipova et al., 2019; Garipova et al., 2021; Gupta et al., 2016; Lima et al., 2012; Shoura et al., 2017; Wang & Xing, 2019; Yesbolatova et al., 2019.

Disclosure statement

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References

- Bashkatov, S. A., & Garipova, M. I. (2022). On the age-specific neurochemical and endocrine biomarkers of temperament traits in adolescents. *Current Opinion in Behavioral Sciences*, 43, 118–124. doi: 10.1016/j.cobeha.2021.09.002.
- Chiellini, G., Nguyen, N.-H., Apriletti, J. W., Baxter, J. D., & Scanlan, T. S. (2002). Synthesis and biological activity of novel thyroid hormone analogues: 5'-aryl substituted GC-1 derivatives. *Bioorganic & Medicinal Chemistry*, 10(2), 333–346. doi: 10.1016/S0968-0896(01)00284-X.
- Garipova, M. I., & Usmanova, R. R. (2013). Isolation and partial characterization of a general hormone transporting blood protein complex. *Journal of Biomolecular Structure and Dynamics*, 31(sup1), 118–118. doi: 10.1080/07391102.2013.786425.
- Garipova, M. I., Fedyayev, V. V., Farkhutdinov, R. G., & Sotnikova, J. M. (2021). Identification of a compound similar to triiodothyronine in the cells of higher plants. *Proceedings of Universities. Applied Chemistry and Biotechnology*, 10(4), 639–646. doi: 10.21285/2227-2925-2020-10-4-639-646.
- Garipova, M. I., Shigapova, A. I., Farkhutdinov, R. G., Fedyayev, V. V., Sotnikova, J. M., & Yakupova, A. B. (2019). The distribution of 3,5,3-triiodothyronine between the transport systems of blood and nuclei of the tissues. *Journal of Biomolecular Structure & Dynamics*, 37(s1), 43–44. doi: 10.1080/07391102.2019.1604468
- Gupta, A., Wamankar, S., Gidwani, B., & Kaur, C. D. (2016). Herbal drugs for thyroid treatment. *International Journal of Pharmacy and Biological Sciences*, 6(1), 62–70.
- Lima, S. T. C., Merrigan, T. L., & Rodrigues, E. D. (2012). Synthetic and plant derived thyroid hormone analogs. In L. S. Ward (Ed.), *Thyroid and parathyroid diseases – New insights into some old and some new issues* (pp. 221–235). In Tech. doi: 10.5772/35134.
- Wang, F., & Xing, J. (2019). Classification of thyroid hormone receptor agonists and antagonists using statistical learning approaches. *Molecular Diversity*, 23(1), 85–92. doi: 10.1007/s11030-018-9857-9.

10. Mass spectrometry-based proteomic analysis of glioblastoma-derived extracellular vesicles and healthy brain cells for biomarkers Identification

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Cellular communication within the brain plays a crucial role in maintaining physiological equilibrium and triggering the onset of diseases. Extracellular vesicles (EVs) are released by donor cells and encapsulate RNA, DNA, and proteins that influence gene and protein expression upon uptake by recipient cells. Transcriptomic and proteomic cell alterations have been associated with the development and progression of glioblastomas. This study aims to identify EV-derived glioblastoma biomarkers through a comparative mass spectrometry-based proteomic analysis of glioblastoma (LN-229) cells and healthy human neurons, astrocytes, and endothelial brain cells (HEBC), therefore, advancing the understanding of the underlying mechanisms of glioblastoma disease progression. EVs were extracted from the cells' conditioned media *via* polymer precipitation. Acid treatment and trypsin digestion prepared the samples for injection. Fifty nanograms of peptides were injected for mass spectrometry analysis. Each experimental group included three biological replicates. Data analysis was performed using Spectronaut software (version 18.3, Biognosys). The statistically significant proteins were annotated using the STRING database. Gene Ontology enrichment analysis was conducted to analyze the biological significance of the overexpressed proteins. The analysis focused on identifying glioblastoma EV-derived proteins that were consistently overexpressed across at least two of the neurons, astrocytes, and HEBC experimental groups. Comparison of EVs derived from glioblastoma and healthy human astrocytes, neurons, and HEBC revealed a subset of 20 proteins that were identified as potential biomarkers of glioblastoma. This study sheds light on the role of EVs in glioblastoma metastasis, and their biomarker's potential for early detection and therapeutic interventions in glioblastoma patients.

11. Structural basis for SARS-CoV-2 spike CD4+ T-cell Epitope dominance

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KEYWORDS

COVID-19; antigen processing

Antigen processing potentially limits priming and/or recall of T cells because epitope peptides may be destroyed by proteolysis or resist extraction from the protein. SARS-CoV-2 infection and vaccination prime spike-specific CD4⁺ T cells with substantially different epitope dominance patterns, and most epitope-clusters exhibit promiscuous dominance with respect to HLA background. Antigen processing likelihood (APL) of epitope peptides has been analyzed on the basis of the 3D structure of the spike trimer. Epitopes that score high in APL generally occur in stable protein segments adjacent to unstable protein segments that serve as early cleavage sites in proteolytic antigen processing. When calculated on the basis of the all-RBD-down conformation of spike, 37% of the vaccine-associated epitopes are well predicted by APL. The known CD4⁺ epitopes were divided into two 9-peptide pools: 9S and 9U, for conformationally stable and unstable regions of spike, respectively. Using PBMC samples collected by the TUCAIN SeroNet Serology Center in the New Orleans area since May 2020, IL-2 Elispots were enumerated upon stimulation with the 9S and 9U pools. Following infection, responses to the 9U pool were more frequent and correlated with pseudovirus neutralization titers in the plasma. Remarkably the bias toward response to the 9U pool was reversed in samples from vaccinated subjects. A modest collection of paired samples from subjects that were infected and later vaccinated also indicated a shift from a low ratio of 9S/9U to high ratio of 9S/9U. Epitope frequencies reported in the Immune Epitope Database confirm the bias toward 9U for convalescents and toward 9S for vaccines. Spike processing pathways and consequences are currently under investigation.

Disclosure statement

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12. Functional studies of the roles of topoisomerases in eccDNA biogenesis and topology

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Extrachromosomal-circular DNAs (eccDNAs) have been found in every organism examined for their presence, including both normal and diseased cells. Using both classical and novel methods that fractionate DNA molecules according to topology, this circular species was shown to consist of molecules ranging in size from ~100bp to >1Mbp. Contrasting with this broad size distribution, next-generation sequencing reveals that eccDNAs originate from a limited repertoire of discrete and conserved linear-genomic loci in normal backgrounds (e.g. wild-type *C. elegans* and human genomes). Our fundamental understanding of eccDNA biogenesis mechanism(s) remains fragmentary. One plausible model connects eccDNA formation with spontaneous double-stranded DNA breaks occurring in regions of high superhelical strain. The study described here addresses how depleted cellular levels of topo-I and topo-II α proteins affect the distribution of eccDNA species in a human cell line. We use an auxin-inducible protein-degradation (degron) system to evaluate the role of topoisomerases in eccDNA formation in HCT-116 cells; specifically, whether depletion of either or both topoisomerase activities alter levels of known eccDNA species and/or uncover additional sites on the linear genome that contribute to the eccDNA repertoire. In addition to generating insights into possible mechanisms of eccDNA formation, this study may have implications for how modulation of topoisomerase activity affects eccDNA distributions in cancer cells treated with topoisomerase-inhibiting chemotherapy drugs. For additional information see Gaubatz, 1990.

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References

- Gaubatz, J. W. (1990). Extrachromosomal circular DNAs and genomic sequence plasticity in eukaryotic cells. *Mutation Research*, 237(5–6), 271–292. doi: [10.1016/0921-8734\(90\)90009-g](https://doi.org/10.1016/0921-8734(90)90009-g).
- Shoura, M. J., Gabdank, I., Hansen, L., Merker, J., Gotlib, J., Levene, S. D., & Fire, A. Z. (2017). Intricate and cell type-specific populations of endogenous circular DNA (eccDNA) in *Caenorhabditis elegans* and *Homo sapiens*. *G3*, 7(10), 3295–3303. doi: [10.1534/g3.117.300141](https://doi.org/10.1534/g3.117.300141).
- Yesbolatova, A., Natsume, T., Hayashi, K., & Kanemaki, M. T. (2019). Generation of conditional auxin-inducible degron (AID) cells and tight control of degron-fused proteins using the degradation inhibitor auxinole. *Methods*, 164–165, 73–80. doi: [10.1016/j.ymeth.2019.04.010](https://doi.org/10.1016/j.ymeth.2019.04.010).

13. Unmasking a hidden DNA-supercoil relaxation activity in a Site-specific recombination system

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Enzymes belonging to the tyrosine superfamily of site-specific recombinases generate knotted products when acting on inverted target sites in circular DNA. For the Cre/loxP system low-complexity torus knots are formed almost exclusively even for highly supercoiled DNA substrates. This is surprising because random-collision models that characterize the recombination reaction predict knot-type distributions of increasing complexity coupled to repeated reaction cycles and the associated release of mechanical energy stored in the supercoiled DNA substrate. We have resolved this puzzle by uncovering an additional hidden DNA-supercoil relaxation activity in Cre/loxP recombination. To this end, we analyzed the time dependence of Cre-dependent topological transitions in supercoiled DNA by using a chemical master-equation model for the time evolution of DNA-substrate topology. We show that the time-dependent knot distributions observed in experiments can only be quantitatively explained by a model that includes

excess DNA unwinding activity, which was also quantified experimentally. Thus, analysis of the dynamic transitions between topological states in knotted supercoiled DNA unravels an important mechanism in Cre/loxP recombination that was not accessible using previous methods. For additional information see Crisona et al., 1999.

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References

- Crisona, N. J., Weinberg, R. L., Peter, B. J., Summers, D. W., & Cozzarelli, N. R. (1999). The topological mechanism of phage lambda integrase. *Journal of Molecular Biology*, 289(4), 747–775. doi: [10.1006/jmbi.1999.2771](https://doi.org/10.1006/jmbi.1999.2771).
- Spengler, S. J., Stasiak, A., & Cozzarelli, N. R. (1985). The stereostructure of knots and catenanes produced by phage integrative recombination: Implications for mechanism and DNA structure. *Cell*, 42(1), 325–334. doi: [10.1016/s0092-8674\(85\)80128-8](https://doi.org/10.1016/s0092-8674(85)80128-8).

14. GPU acceleration of conformational stability computation for CD4⁺ T cell epitope prediction

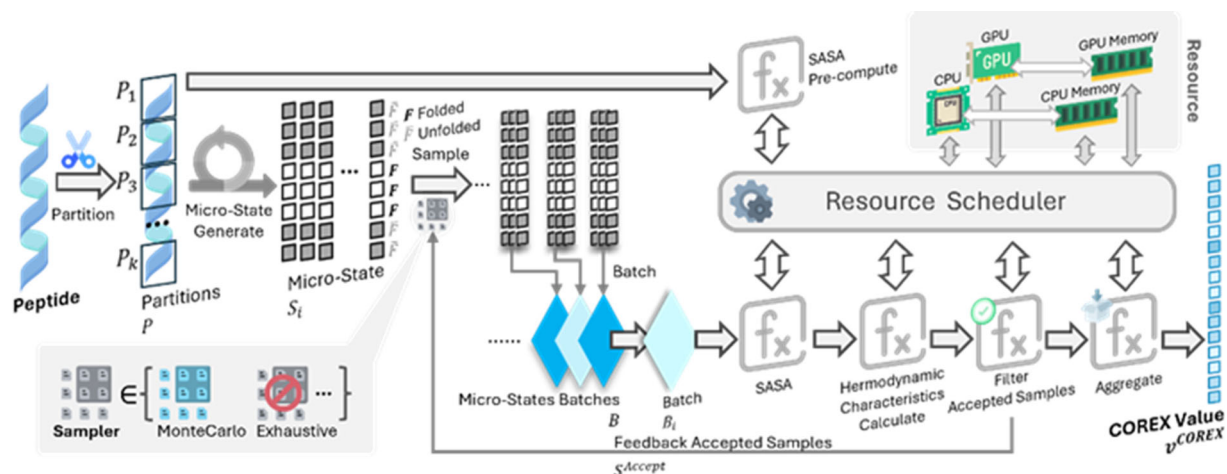
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KEYWORDS

Computational epitope prediction; CD4⁺ T cell response; antigen processing; free energy calculation; protein structure; graphics processing unit; high performance computation



CD4⁺ T cells play a crucial role in adaptive immunity and are a significant component of immunological response in many settings. Computational prediction of which antigenic peptides are presented and bind to T cells is a problem that has been studied for several decades. Current efforts apply supervised learning methods to predict peptide-MHCII binding but do not incorporate the role of antigen processing. To address this, our group developed the Antigen Processing Likelihood (APL) algorithm, which relies on a free energy-based conformational stability metric known as 'COREX'. However, COREX requires the analysis of a potentially large conformational ensemble and is thus computationally intensive. In prior work, we parallelized the original COREX algorithm on CPU cores and reduced the computation time from hours to minutes. In our current work, we have further accelerated COREX utilizing many thousands of GPU cores and have brought computation times down from minutes to seconds. To do this, we developed a novel memorization technique to share common energy terms between conformations and developed a novel Monte Carlo sampling approach. We demonstrate the effectiveness of our approach, both in terms of computation time and quality of predictions, on several benchmark sets of antigens. For additional information see Hilser & Freire, 1996; Mettu et al., 2016.

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References

- Bhattacharya, A., Wrabl, J. O., Landry, S. J., et al. (2023). Parallel computation of conformational stability for CD4⁺ T-cell epitope prediction. 2023 IEEE International Conference on Bioinformatics and Biomedicine (pp. 88–93). doi: [10.1109/BIBM58861.2023.10385333](https://doi.org/10.1109/BIBM58861.2023.10385333).
- Hilser, V. J., & Freire, E. (1996). Structure-based calculation of the equilibrium folding pathway of proteins. Correlation with hydrogen exchange protection factors. *Journal of Molecular Biology*, 262(5), 756–772. doi: [10.1006/jmbi.1996.0550](https://doi.org/10.1006/jmbi.1996.0550).
- Mettu, R. R., Charles, T., & Landry, S. J. (2016). CD4⁺ T-cell epitope prediction using antigen processing constraints. *Journal of Immunological Methods*, 432, 72–81. doi: [10.1016/j.jim.2016.02.013](https://doi.org/10.1016/j.jim.2016.02.013).

15. Hi-BDiSCO: folding 3D mesoscale genome structures from Hi-C data using Brownian dynamics

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KEYWORDS

Hi-C; mesoscale model; chromatin; 3D reconstruction; Brownian dynamics

The structure and dynamics of the eukaryotic genome are intimately linked to gene regulation and transcriptional activity. Many chromosome conformation capture experiments like Hi-C have been developed to detect genome-wide contact frequencies and quantify loop/compartments structures for different cellular contexts and time-dependent processes. However, a full understanding of these events requires explicit descriptions of representative chromatin and chromosome configurations. With the exponentially growing amount of data from Hi-C experiments, many methods for deriving 3D structures from contact frequency data have been developed. Yet, most reconstruction methods use polymer models with low resolution to predict overall genome structure. Here we present a Brownian Dynamics (BD) approach termed Hi-BDiSCO for producing 3D genome structures from Hi-C and Micro-C data using our mesoscale-resolution chromatin model based on the Discrete Surface Charge Optimization (DiSCO) model. Our approach integrates reconstruction with chromatin simulations at nucleosome resolution with appropriate biophysical parameters. Following a description of our protocol, we present applications to the NXN, HOXC, HOXA, and Fbn2 mouse genes ranging in size from 50 to 100 kb. Such nucleosome-resolution genome structures pave the way for pursuing many biomedical applications related to the epigenomic regulation of chromatin and control of human disease. For additional information see Li & Schlick, 2023; Li et al., 2022; Li et al., 2023.

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Disclosure statement

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References

- Li, Z., & Schlick, T. (2023). Hi-BDiSCO: Folding 3D mesoscale genome structures from Hi-C data using Brownian dynamics. *Nucleic Acids Research*, 52(2), 583–599. doi: [10.1093/nar/gkad1121](https://doi.org/10.1093/nar/gkad1121).
- Li, Z., Portillo-Ledesma, S., & Schlick, T. (2022). Brownian dynamics simulations of mesoscale chromatin fibers. *Biophysical Journal*, 122(14), 2884–2897. doi: [10.1016/j.bpj.2022.09.013](https://doi.org/10.1016/j.bpj.2022.09.013).

Li, Z., Portillo-Ledesma, S., & Schlick, T. (2023). Techniques for and challenges in reconstructing 3D genome structures from 2D chromosome conformation capture data. *Current Opinion in Cell Biology*, 83, 102209. doi: 10.1016/j.ceb.2023.102209.

16. Implications of stoichiometric compositions on order and disorder in natural proteins

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KEYWORDS

Protein folding; stoichiometry; protein structure; protein function; proteins; evolution

More than a decade ago, rigorous analyses of structural data of thousands of naturally occurring folded proteins yielded a ‘margin of life’ for stoichiometric composition, i.e. percentage occurrence of individual amino acids, of protein sequences (Mittal et al., 2010; Mittal & Jayaram, 2011a, 2011b). This ‘margin of life’ refers to the lower-than-expected variances in percentage occurrence of individual amino acids in protein sequences (Mezei, 2011). Subsequently, the constraints on stoichiometric compositions have been confirmed over a large sequence space for almost all known protein sequences (even in absence of structural data). While the earlier work was based on the largest structural dataset at that time (several thousands of structures), in this work further explorations on compositional considerations for known protein sequences are discussed. The relationships between occurrences of all possible di-, tri-, tetra-, and penta- peptides, in more than half-a-million curated (manually annotated and reviewed) primary sequences, with their various physico-chemical properties and individual proportions of amino acids are explored (Mittal, Changani, & Taparia, 2021; Mittal, Changani, Taparia, et al., 2021; Mittal et al., 2020). A key result conclusively shows that stoichiometric constraints on amino acids limit the primary sequence space of proteins in nature rather than any perceived limitations of evolutionary sampling of the primary sequence space for natural occurrence of proteins. Thus, while having profound evolutionary implications (Mittal & Chauhan, 2022) toward our insights into occurrence of naturally occurring protein sequences, the results discussed also promise to serve as a guide for creating stable and structurally controlled and/or ‘disordered’ designer proteins.

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Disclosure statement

No potential conflict of interest was reported by the author(s).

References

- Mezei, M. (2011). Discriminatory power of stoichiometry-driven protein folding? *Journal of Biomolecular Structure & Dynamics*, 28(4), 625–626. doi: 10.1080/073911011010524966.
- Mittal, A., & Chauhan, A. (2022). Aspects of biological replication and evolution independent of the central dogma: Insights from protein-free vesicular transformations and protein-mediated membrane remodeling. *The Journal of Membrane Biology*, 255(2–3), 185–209. doi: 10.1007/s00232-022-00230-4.
- Mittal, A., & Jayaram, B. (2011a). Backbones of folded proteins reveal novel invariant amino acid neighborhoods. *Journal of Biomolecular Structure & Dynamics*, 28(4), 443–454. doi: 10.1080/073911011010524954.
- Mittal, A., & Jayaram, B. (2011b). The newest view on protein folding: Stoichiometric and spatial unity in structural and functional diversity. *Journal of Biomolecular Structure and Dynamics*, 28(4), 669–674. doi: 10.1080/073911011010524984.
- Mittal, A., Changani, A. M., & Taparia, S. (2020). What limits the primary sequence space of natural proteins? *Journal of Biomolecular Structure & Dynamics*, 38(15), 4579–4583. doi: 10.1080/07391102.2019.1682051.
- Mittal, A., Changani, A. M., & Taparia, S. (2021). Unique and exclusive peptide signatures directly identify intrinsically disordered proteins from sequences without structural information. *Journal of Biomolecular Structure & Dynamics*, 39(8), 2885–2893. doi: 10.1080/07391102.2020.1756410.
- Mittal, A., Changani, A. M., Taparia, S., Goel, D., Parihar, A., & Singh, I. (2021). Structural disorder originates beyond narrow stoichiometric margins of amino acids in naturally occurring folded proteins. *Journal of Biomolecular Structure & Dynamics*, 39(7), 2364–2375. doi: 10.1080/07391102.2020.1751299.
- Mittal, A., Jayaram, B., Shenoy, S. R., & Bawa, T. S. (2010). A stoichiometry driven universal spatial organization of backbones of folded proteins: Are there Chargaff’s rules for protein folding? *Journal of Biomolecular Structure & Dynamics*, 28(2), 133–142. doi: 10.1080/07391102.2010.10507349.

17. Deriving polymer properties of chromatin from nucleosome-resolution contact map data

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KEYWORDS

Chromatin; coarse-grained models; Micro-C

It is common to model chromatin as a bead spring chain. However, the polymer properties of the chromatin—its stretching elasticity, bending elasticity, the nature of inter-bead interactions, etc.—are not well understood (Figure 1). To bridge this gap, we simulated a large ensemble of chromatin configurations at near-nucleosome (200 bp) resolution, consistent with recently published contact map data. We then systematically coarse-grained the configurations and predicted quantities essential for the polymer representation of chromatin. Unlike the prevalent notion, we show that

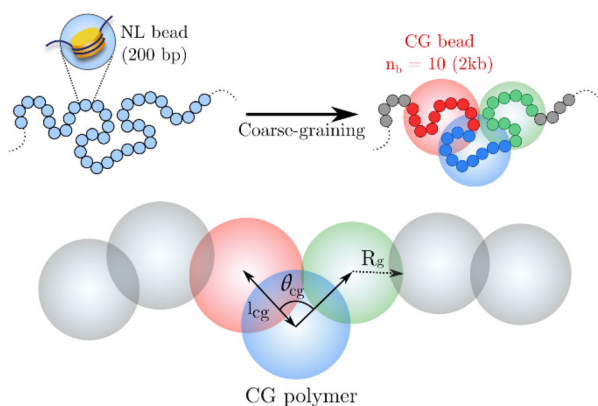


Figure 1. Schematic representing the systematic coarse-graining of nucleosome resolution chromatin conformations.

coarse-grained chromatin polymer beads should be considered soft particles that can overlap, and we derive an intrachromatin soft potential. We show that accounting for such softness is crucial to predicting 3D distances from simulations in a self-consistent manner. For additional information see Hsieh et al., 2020; Kadam et al., 2023.

Disclosure statement

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References

- Hsieh, T.-H. S., Cattoglio, C., Slobodyanyuk, E., Hansen, A. S., Rando, O. J., Tjian, R., & Darzacq, X. (2020). Resolving the 3D landscape of transcription-linked mammalian chromatin folding. *Molecular Cell*, 78(3), 539–553.e8. doi: 10.1016/j.molcel.2020.03.002.
- Kadam, S., Kumari, K., Manivannan, V., Dutta, S., Mitra, M. K., & Padinhateeri, R. (2023). Predicting scale-dependent chromatin polymer properties from systematic coarse-graining. *Nature Communications*, 14(1), 4108. doi: 10.1038/s41467-023-39907-2.

18. [In absentia] unveiling the anti-oncogenic effect of rosmarinic acid (RosA) on hepatocellular carcinoma: an integrated network pharmacology, molecular docking, and dynamics approach

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KEYWORDS

Hepatocellular carcinoma; rosmarinic acid (RosA); network pharmacology; molecular docking and molecular dynamics simulation

One of the extremely complicated oncology hepatocellular carcinoma (HCC) upsurges the occurrences and mortalities with 906,000 fresh incidences annually at the global index. The third most rampant source of cancer-associated fatalities poses a challenge for scientists and hepatologists across the world. The therapeutic plant derived phytoconstituent rosmarinic acid (RosA) from *Rosmarinus officinalis* L. has emerged as a new therapeutic opportunities with less toxicity and is established to exhibit anti-microbial, anti-viral, anti-oxidant, anti-proliferative, and anti-inflammatory activities. However, the underlying molecular mechanism for anti-oncogenic effects of Rosmarinic acid over HCC-associated targets through network pharmacology still remains uncultivated. The present study attempts to analyze the anti-HCC potential of RosA through an assimilated approach of network pharmacology. Herein, numerous open-access databases were used to retrieve HCC-associated and RosA-associated targets leading to identification of a total of 104 potential overlapping targets among HCC and RosA. The 104 potential targets were further subjected to gene ontology, gene-disease association, and Pathway enrichment analysis by employing EnrichR webserver. Subsequently, Protein-protein interaction network was constructed by Cytoscape software, and the top 10 hub nodes were identified through implementing CytoNCA plugin. Consequently, the top 10 significant hub nodes were validated by survival analysis and expression analysis through adapting KM plotter and GEPIA2 databases, respectively. Accordingly, the drug-like and ADME properties of RosA were evaluated by QikProp tool available with Maestro module of Schrödinger software. Additionally, the findings from Molecular Docking and Molecular dynamic simulations performed by Glide module and Desmond module (Schrodinger software) respectively, indicate ESR1, EGFR, MMP9, and PPARG as potential anti-HCC RosA related targets with promising complex stability. Sequentially, the outcomes from free energy of binding of RosA to protein targets computed with the Prime module of the Schrödinger suite employing the MM/GBSA (Molecular Mechanics/Generalized Born Surface Area) procedure revealed that ESR1 and RosA have high binding affinity. Our findings not only reveal the anti-oncogenic role of RosA but also provide novel insights illuminating the identified target ESR1 as scientific foundation for anti-oncogenic clinical application of RosA in HCC therapeutics. For additional information see Andrade et al., 2018; Hopkins, 2008.

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Government of Gujarat for providing BIN-Node Facility to our department.

Disclosure statement

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References

- Andrade, J. M., Faustino, C., Garcia, C., Ladeiras, D., Reis, C. P., & Rijo, P. (2018). *Rosmarinus officinalis* L.: An update review of its phytochemistry and biological activity. *Future Science OA*, 4(4), FSO283. doi: 10.4155/fsoa-2017-0124.
- Dixit, N., Motwani, H., Patel, S. K., Rawal, R. M., & Solanki, H. A. (2023). Decoding the mechanism of andrographolide to combat hepatocellular carcinoma: A network pharmacology integrated molecular docking and dynamics approach. *Journal of Biomolecular Structure & Dynamics* 1–19. doi: 10.1080/07391102.2023.2256866.
- Hopkins, A. L. (2008). Network pharmacology: The next paradigm in drug discovery. *Nature Chemical Biology*, 4(11), 682–690. doi: 10.1038/nchembio.118.

19. Dietary epigenetic modulators: unravelling the cross-kingdom regulation of plant miRNAs in nutrition and disease

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KEYWORDS

Plant microRNAs; cross-kingdom analysis; human gene regulation

MicroRNAs are delivered to host cells through oral consumption and play an important function in the control of host genes. Numerous studies have extensively established the cross-species post-transcriptional regulation potential of plant-derived small non-coding microRNAs (miRNAs). The present study aims to uncover the potential regulatory role of miRNAs derived from *Andrographis paniculata*, *Cyperus rotundus*, *Anethum foeniculum*, *Holarrhena pubescens*, *Picrorhiza kurroa*, *Ocimum basilicum*, *Carica papaya*, and *Bacopa monnieri*. Using computational tools and bioinformatics approach, resulted in identification of known and putative novel miRNAs regulating human genes involved in various diseases. The putative miRNAs were predicted by homology search using the Blast algorithm against the miRbase database. The respective human targets were predicted by psRNATarget webserver which were subsequently annotated for gene ontology analysis and pathway enrichment analysis to identify their biological functions and associated diseases. The identified hub

proteins HGF, NRP2, and CCND1 against *A. paniculata* miRNAs show their significant involvement in cancerous pathways including STAT3, MAPK, TGF- β , mTOR, and VEGFR pathways. Deregulation of these pathways has been reported as oncogenic response involved in Hepatocellular Carcinoma, cell proliferation, and metastasis. Interestingly, the found putative novel miRNAs apa-miR-5, apa-miR-1, apa-miR-26, and apa-miR-30 target host genes involved in p38-MAPK, AKT, AMPK, NF- κ B, ERK, WNT signaling, MYD88-dependent cascade, and pathways in cancer leading to Alzheimer's, non-alcoholic fatty liver, alcoholic liver diseases and hepatocellular carcinoma (HCC). Human target genes related to signaling pathways such NF- κ B and MAPK, with TRAF2, CBX1, IL1B, ITGA4, and ITGB1BP1 being the top five hub nodes against *B. monnieri* miRNAs. The novel *C. papaya* miRNAs targets involved in cancer, diabetes, mental illness, and platelet disorder. Similarly, *Cyperus rotundus* microRNAs target genes were involved in high blood pressure, angina pectoris, dysarthria, extrapyramidal disorders, and muscle hypotonia. Our findings postulate novel interpretations regarding modulation of human transcripts by *A. paniculata*, *C. rotundus*, *A. foeniculum*, *H. pubescens*, *P. kurroa*, *O. basilicum*, *C. papaya*, and *B. monnieri* miRNAs and exhibit the regulation of human diseases by these plant-derived miRNAs. Though our study elucidates miRNAs as novel therapeutic agents, however experimental validations for assessment of therapeutic potential of these miRNAs are still warranted. For additional information see Jha et al., 2022; Motwani et al., 2023; Trivedi et al., 2023.

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Disclosure statement

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References

- Jha, N., Mangukia, N., Gadhavi, H., Patel, M., Bhavsar, M., Rawal, R., & Patel, S. K. (2022). Small RNA sequencing and identification of papaya (*Carica papaya* L.) miRNAs with potential cross-kingdom human gene targets. *Molecular Genetics and Genomics*, 297(4), 981–997. doi: 10.1007/s00438-022-01904-3.
- Motwani, H., Patel, M., Nanavaty, V., Dixit, N., Rawal, R. M., Patel, S. K., & Solanki, H. A. (2023). Small RNA sequencing and identification of *Andrographis paniculata* miRNAs with potential cross-kingdom human gene targets. *Functional & Integrative Genomics*, 23(1), 55. doi: 10.1007/s10142-023-00976-7.
- Trivedi, T. S., Patel, M. P., Nanavaty, V., Mankad, A. U., Rawal, R. M., & Patel, S. K. (2023). MicroRNAs from *Holarrhena pubescens* stems: Identification by small RNA sequencing and their potential contribution to human gene targets. *Functional & Integrative Genomics*, 23(2), 149. doi: 10.1007/s10142-023-01078-0.

20. Disorder, function, and phase separation

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Disorder creates function in many biomolecular systems, including nucleic acids and proteins. Here, the properties of peptides and proteins in water at varying concentrations are used to understand the structural, thermodynamic, and kinetic changes that occur during a simulated liquid-liquid phase separation. Thermodynamic signatures of the aggregation of peptides are remarkably similar to the thermodynamics of folding or collapse of longer polymers in water. Disordered protein sequences are found to be preferentially enhanced in intracellular condensates. We decompose the free energy to study the entropic contributions. Enthalpy-entropy compensation between the components in the phase separated systems gives mechanistic hypotheses for the driving forces. Dynamic turn over in condensates determines the kinetics of signaling in many experimentally examined systems. We simulated the diffusion constant for peptides in aqueous solutions. Both sequence and peptide conformation were found to influence diffusion in these systems with sequence dominating the magnitude of the differences. We found the most compact structures for each sequence diffused the fastest in the peptide rich phase.

21. Implicit modeling of protein binding to chromatin fibers

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KEYWORDS

Chromatin; protein binding; mesoscale modeling

Genome expression is regulated by the binding of different proteins, such as transcription factors (TFs), repressors, and remodelers. How such protein binding affects chromatin architecture at the nucleosome level remains not fully understood. In this work, we illustrate how TF binding can be introduced implicitly into our mesoscale chromatin model to obtain insights into TF mechanisms of actions. In particular, we introduce TF binding by harmonic constraints and by bending and opening of the linker DNA. Overall, our results show that although our implicit modeling of TF binding are

simple strategies, they provide clear trends and guiding insights into the regulation of chromatin architecture by protein binding.

Disclosure statement

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22. Cyclization-based measurements of DNA bending flexibility at eccDNA hotspots

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Extrachromosomal-circular DNAs (eccDNAs) are a class of endogenous circular-DNA elements in eukaryotic cells (Liao et al., 2020). These DNA circles are derived from sequences on the linear genome and can contribute to genetic diversity (Shoura et al., 2017). eccDNAs have been found in every eukaryotic organism studied thus far, including humans, and are present in both normal and diseased cells (Chen et al., 2022). Although the biogenesis mechanisms of eccDNA remain unclear, these circular sequence elements map to discrete and conserved regions within the eukaryotic domain. Some sequences are more commonly found within the eccDNA population than others, however, no comprehensive model has yet been advanced that explains the roles of sequence and DNA bending flexibility in eccDNA formation.

In this study, we investigate the role of DNA sequence in driving eccDNA formation. We employ cyclization *via* Cre/loxP site-specific recombination to investigate the bending flexibility eccDNA-sequence mimics to elucidate the relationship between DNA sequence and propensity of ring closure as quantitated by the Jacobson–Stockmayer factor (J factor) (Shoura et al., 2012, 2020). J factor values for a series of eccDNA analogs and scrambled controls are obtained from time-dependent Förster Resonance Energy Transfer (FRET) measurements involving fluorescent donor and acceptor molecules positioned at the opposite ends of the DNA fragment (Shoura et al., 2012). By comparing the J factors of eccDNA analogs to those of their corresponding

controls, we will investigate the thermodynamics of DNA looping as a function of eccDNA sequence, which may offer valuable insights into the mechanisms underlying eccDNA biogenesis (Crothers et al., 1992).

Disclosure statement

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References

- Chen, Z., Qi, Y., He, J., Xu, C., Ge, Q., Zhuo, W., Si, J., & Chen, S. (2022). Distribution and characterization of extrachromosomal circular DNA in colorectal cancer. *Molecular Biomedicine*, 3(1), 38. doi: [10.1186/s43556-022-00104-0](https://doi.org/10.1186/s43556-022-00104-0).
- Crothers, D. M., Drak, J., Kahn, J. D., & Levene, S. D. (1992). DNA bending, flexibility, and helical repeat by cyclization kinetics. *Methods in Enzymology*, 212, 3–29. doi: [10.1016/0076-6879\(92\)12003-9](https://doi.org/10.1016/0076-6879(92)12003-9).
- Liao, Z., Jiang, W., Ye, L., Li, T., Yu, X., & Liu, L. (2020). Classification of extrachromosomal circular DNA with a focus on the role of extrachromosomal DNA (eccDNA) in tumor heterogeneity and progression. *Biochimica et Biophysica Acta*, 1874(1), 188392. doi: [10.1016/j.bbcan.2020.188392](https://doi.org/10.1016/j.bbcan.2020.188392).
- Shoura, M. J., Gabdank, I., Hansen, L., Merker, J., Gotlib, J., Levene, S. D., & Fire, A. Z. (2017). Intricate and cell type-specific populations of endogenous circular DNA (eccDNA) in *Caenorhabditis elegans* and *Homo sapiens*. *Genes Genomes Genetics*, 7(10), 3295–3303. doi: [10.1534/g3.117.300141](https://doi.org/10.1534/g3.117.300141)
- Shoura, M. J., Giovan, S. M., Vetcher, A. A., Ziraldo, R., Hanke, A., & Levene, S. D. (2020). Loop-closure kinetics reveal a stable, right-handed DNA intermediate in Cre recombination. *Nucleic Acids Research*, 48(8), 4371–4381. doi: [10.1093/nar/gkaa153](https://doi.org/10.1093/nar/gkaa153).
- Shoura, M. J., Vetcher, A. A., Giovan, S. M., Bardai, F., Bharadwaj, A., Kesinger, M. R., & Levene, S. D. (2012). Measurements of DNA-loop formation via Cre-mediated recombination. *Nucleic Acids Research*, 40(15), 7452–7464. doi: [10.1093/nar/gks430](https://doi.org/10.1093/nar/gks430).

23. Transitions between SARS-CoV-2 RNA pseudoknots shed insight on viral mechanism

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KEYWORDS

Transition path sampling; RNA; frameshifting; pseudoknot

Coronaviruses employ programmed ribosomal frameshifting to produce viral proteins from a compact viral genome. A 3-stem RNA pseudoknot has been identified to stimulate frameshifting for SARS-CoV-2. We characterized an alternative pseudoknot using our RNA-As-Graphs framework coupled with chemical reactivity experiments. Although structural plasticity has been proposed to play key roles in frameshifting, transition paths among different SARS-CoV-2 FSE have yet to be reported. Here, we delineate transition pathways between two key FSE pseudoknots by transition path sampling (TPS) simulations and reveal a heterogeneous, multi-hub conformational landscape with complex multiple transition paths. This transition critically tunes the tension through the spacer region, whose unwinding directs the placement of the viral RNA in the narrow mRNA channel. Our work further shows how the alternative pseudoknot affects bound ribosome conformations and why the experimentally captured 3-stem pseudoknot is preferred for frameshifting. This success of capturing an unprecedented large-scale RNA 2D structure transition along a highly complex path ensemble highlights the inherent multifarious aspects that viruses develop for virulence and survival. This work enhances our understanding of viral frameshifting and provides new insights to target key transitions for therapeutic applications. For additional information see Bhatt et al., 2021; Bolhuis et al., 1998; Schlick et al., 2021.

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References

- Bhatt, P. R., Scaiola, A., Loughran, G., Leibundgut, M., Kratzel, A., Meurs, R., Dreos, R., O'Connor, K. M., McMillan, A., Bode, J. W., Thiel, V., Gatfield, D., Atkins, J. F., & Ban, N. (2021). Structural basis of ribosomal frameshifting during translation of the SARS-CoV-2 RNA genome. *Science*, 372(6548), 1306–1313. doi: [10.1126/science.abf3546](https://doi.org/10.1126/science.abf3546).
- Bolhuis, P. G., Dellago, C., & Chandler, D. (1998). Sampling ensembles of deterministic transition pathways. *Faraday Discussions*, 110, 421–436. doi: [10.1039/A801266K](https://doi.org/10.1039/A801266K).
- Schlick, T., Zhu, Q., Dey, A., Jain, S., Yan, S., & Laederach, A. (2021). To knot or not to knot: Multiple conformations of the SARS-CoV-2 frame-shifting RNA element. *Journal of the American Chemical Society*, 143(30), 11404–11422. doi: [10.1021/jacs.1c03003](https://doi.org/10.1021/jacs.1c03003).

24. The nucleosome reference frame and standard geometries for octasomes

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KEYWORDS

DNA; nucleosome; nucleosome reference frame; standard geometries

Nucleosomes are the building blocks of eukaryotic genomes and thus fundamental to all genetic processes (Figure 1). There are over 533 nucleosome structures in the Research Collaboratory for Structural Bioinformatics (RCSB) (Berman et al., 2000; Burley et al., 2020). Collectively numerous variants and species are present, as are, sub-nucleosomal and super-nucleosomal assemblies within the nucleosome family. The relative orientation and location of the histones with respect to each other are highly conserved in all standard octasomes containing 145, 146, or 147 base pairs of DNA. This observation is used to establish a nucleosome reference frame that enables us to describe and compare the gross structure and organization of all of them. We observe that the cumulative sums of the DNA structural parameters Twist, Rise, and arc length, as a function of base pair step, are linear with R2 values exceeding 0.999 for almost all octasome structures. These observations inform our comparison of the location and orientation of the DNA director frames observed

in X-ray structures to ideal superhelix values. The comparisons reveal a regular variation in the superhelix radius and a distinct straightening of the superhelix over the outer most turn of the DNA helix. Once aligned, the path of DNA over the histone core exhibits conserved, single-track, and multipath regions. To demonstrate the utility of the proposed reference geometries, standard and distorted octasome structures, super-nucleosomal structures, nucleosomes with linker DNA, and nucleosomes in closed circular DNA are analyzed.

Disclosure statement

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References

- Berman, H. M., Westbrook, J., Feng, Z., Gilliland, G., Bhat, T. N., Weissig, H., Shindyalov, I. N., & Bourne, P. E. (2000). The protein data bank. *Nucleic Acids Research*, 28(1), 235–242. doi: [10.1093/nar/28.1.235](https://doi.org/10.1093/nar/28.1.235).
- Burley, S. K., Bhikadiya, C., Bi, C., Bittrich, S., Chen, L., Crichlow, G. V., Christie, C. H., Dalenberg, K., Di Costanzo, L., Duarte, J. M., Dutta, S., Feng, Z., Ganesan, S., Goodsell, D. S., Ghosh, S., Green, R. K., Guranović, V., Guzenko, D., Hudson, B. P., ... Zhuravleva, M. (2020). RCSB protein data bank: Powerful new tools for exploring 3D structures of biological macromolecules for basic and applied research and education in fundamental biology, biomedicine, biotechnology, bioengineering and energy sciences. *Nucleic Acids Research*, 49(D1), D437–D451. doi: [10.1093/nar/gkaa1038](https://doi.org/10.1093/nar/gkaa1038).

25. Single-molecule size and topological characterization of circular DNA

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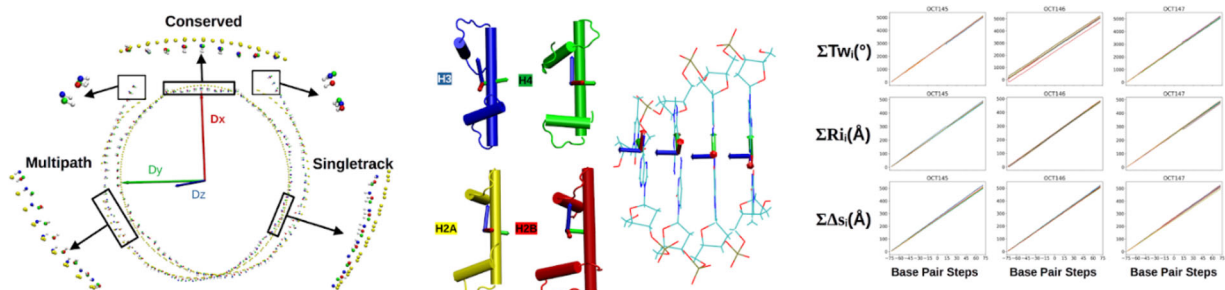


Figure 1. Left to right: DNA pathways around the histone core, histone, and DNA director frames, and the cumulative sums of various DNA structural parameters: twist, rise, and arc length.

KEYWORDS

Single-molecule; eccDNA; solid-state nanopore; sequencing

Extrachromosomal-circular DNA (eccDNA) is a heterogeneous class of endogenous eukaryotic circular DNAs that are variable with respect to both size and sequence. In human cells and other organisms, a subclass of this species is associated with genomic instability and pathologies, such as cancer (Yi et al., 2022). In normal cells, eccDNA populations are typically limited to molecules 200 bp to 20 kbp in size and contain genetic fragments, such as an exon or a single gene. In contrast, tumor-cell eccDNA populations can harbor multiple gene fragments, gene fusions, and potentially amplified copies of complete oncogenes, with circle sizes exceeding 1 Mbp. The diversity of eccDNA sizes presents a challenge to both short-read (Illumina) and long-read [Oxford Nanopore Technology (ONT)] sequencing technologies (Guiblet et al., 2018). Thus, characterizing distributions of molecular size and topology (circular vs. linear) for isolated eccDNA samples before sequencing-library preparation would provide valuable information for circle-sequence assembly. Single-molecule nanopore electrophoresis (SMNE) complements both short- and long-read sequencing as it can provide topological and molecular size distributions based on the magnitude of the nanopore current-blockade signal and translocation time of DNA through the nanopore. Here we present data demonstrating discrimination of size and topology of synthetic dsDNA circles using SMNE. This project will apply SMNE to characterize molecular size and topology distributions in enriched eccDNA populations prepared from human cell lines.

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References

Guiblet, W. M., Cremona, M. A., Cechova, M., Harris, R. S., Kejnovska, I., Kejnovsky, E., Eckert, K., Chiaromonte, F., & Makova, K. D. (2018). Long-read sequencing technology indicates genome-wide effects of non-B DNA on polymerization speed and error rate. *Genome Research*, 28(12), 1767–1778. doi: [10.1101/gr.241257.118](https://doi.org/10.1101/gr.241257.118).

Yi, E., Chamorro González, R., Henssen, A. G., & Verhaak, R. G. W. (2022). Extrachromosomal DNA amplifications in cancer. *Nature Reviews. Genetics*, 23(12), 760–771. doi: [10.1038/s41576-022-00521-5](https://doi.org/10.1038/s41576-022-00521-5).

26. Finding a few allosteric networks: a tale of two cities

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KEYWORDS

p53 tumor suppressor protein; allosteric signaling; molecular dynamics simulation; networks

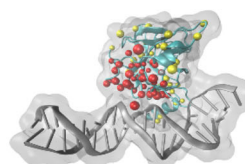
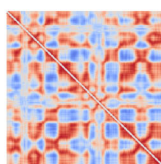
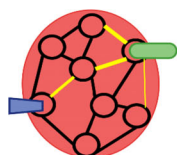
Bridging molecular biophysics and computer science presents significant potential for elucidating complex biological mechanisms and developing novel therapeutic strategies to address currently undruggable diseases, such as cancer. Allosteric signaling in proteins, wherein a small molecule binds to one site and induces a conformational or activity change at a distant site, represents a key area for interdisciplinary study. With this in mind, proteins can be cast as a complex network of interacting residues capable of communicating not only with other residues in spatial proximity but also across the protein via long-range forces. Although this phenomenon has been widely known as a pervasive phenomenon throughout biology first observed in experiment over 50 years ago (Monod et al., 1963), a conceptual framework of how it arises is lacking. Our work focuses on the tumor suppressor protein p53 (Bauer et al., 2016), important not only as an allosteric protein but also as a key target for curing cancers. Combining molecular simulations, network analysis, and machine learning (Aggarwal et al., 2021; Salomon-Ferrer et al., 2013), we explore the dynamics of long range allosteric signaling. Leveraging Artificial Intelligence, we extract information to reverse engineer small molecules to allosterically control proteins to reverse the adverse effects of mutations, paving the way for curing cancer. These new approaches may provide transferrable approaches for inroads for other currently undruggable systems.

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References

- Aggarwal, A., Mittal, M., & Battineni, G. (2021). Generative adversarial network: An overview of theory and applications. *International Journal of Information Management Data Insights*, 1(1), 100004. doi: [10.1016/j.jjimei.2020.100004](https://doi.org/10.1016/j.jjimei.2020.100004).
- Bauer, M. R., Joerger, A. C., & Fersht, A. R. (2016). 2-Sulfonylpyrimidines: Mild alkylating agents with anticancer activity toward P53-compromised cells. *Proceedings of the National Academy of Sciences of the United States of America*, 113(36), E5271–E5280. doi: [10.1073/pnas.1610421113](https://doi.org/10.1073/pnas.1610421113).
- Monod, J., Changeux, J.-P., & Jacob, F. (1963). Allosteric proteins and cellular control systems. *Journal of Molecular Biology*, 6(4), 306–329. doi: [10.1016/S0022-2836\(63\)80091-1](https://doi.org/10.1016/S0022-2836(63)80091-1)
- Salomon-Ferrer, R., Case, D. A., & Walker, R. C. (2013). An overview of the amber biomolecular simulation package. *WIREs Computational Molecular Science*, 3(2), 198–210. doi: [10.1002/wcms.1121](https://doi.org/10.1002/wcms.1121).

27. Probing the mechanisms of epigenetic regulation with molecular simulations

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KEYWORDS

Chromatin structure; epigenetic mechanisms; molecular dynamics

The molecular mechanisms underlying gene regulation rely on complex interactions between DNA, histones, transcription factors, and other effector molecules in the nucleus. The central processes for many epigenetic mechanisms rely on molecules binding to and modifying the nucleosome's structure: a complex of approximately equal parts DNA and histone proteins. Although, in many cases, the structural effects of these modifications are beginning to be well characterized, how they take advantage of and tune nucleosome dynamics is far less well understood. In this talk, I will discuss work in our group that utilizes molecular dynamics simulations to explore the effects of nucleosome modification at atomic-scale resolution. In particular, I will discuss how the introduction of a large ubiquitin moiety can alter DNA compaction and nucleosome formation and how viral nucleosome-like particles create destabilized chromatin structures. Overall, results will show how modifications can alter the structure and dynamics of the nucleosome as a means for controlling gene expression.

28. 'Cracking' the secondary codes that control access to the primary code of DNA

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KEYWORDS

DNA supercoiling; DNA looping; DNA counterions; DNA acting enzymes

Supercoiled and looped DNA surrounded by counterions (from AI Image Generator).

By regulating access to the primary code, supercoiling and looping can be thought of as secondary DNA codes. Toward 'cracking' these secondary codes, we previously discovered that supercoiling and loop length-dependent site-specific base pair disruptions (Fogg 2021) facilitate very sharp bending to allow DNA to adopt the unexpected conformations visualized by cryo-electron tomography (Irobalieva et al., 2015). Our previous molecular dynamics simulations revealed that one flipped base, with enough negative supercoiling, expands into a series of adjacent flipped bases to form denaturation bubbles (Randall et al., 2009). We verified these simulations and further discovered that site-specific base pair disruptions in negatively supercoiled minicircles at one site cause site-specific base pair disruptions at distant sites, a remarkable 'action at a distance' with biological consequences, including capturing DNA gyrase for relaxation (Vayssières et al., 2024). In this talk, I will discuss our new findings demonstrating that cations, most particularly divalent cations, should also be considered a secondary code because they dramatically affect the interplay of negative (but not positive) supercoiling and looping-dependent site-specific base-pair disruptions and DNA shape to regulate DNA activity. For additional information see Vayssières et al., 2024.

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References

- Fogg, J. M., Judge, A. K., Stricker, E., Chan, H. L., & Zechiedrich, L. (2021). Supercoiling and looping promote DNA base accessibility and coordination among distant sites. *Nature Communications*, 12(1), 5683. doi: [10.1038/s41467-021-25936-2](https://doi.org/10.1038/s41467-021-25936-2).

- Irobalieva, R. N., Fogg, J. M., Catanese, D. J., Sutthibutpong, T., Chen, M., Barker, A. K., Ludtke, S. J., Harris, S. A., Schmid, M. F., Chiu, W., & Zechiedrich, L. (2015). Structural diversity of supercoiled DNA. *Nature Communications*, 6(1), 8440. doi: [10.1038/ncomms9440](https://doi.org/10.1038/ncomms9440).
- Randall, G. L., Zechiedrich, L., & Pettitt, B. M. (2009). In the absence of writhe, DNA relieves torsional stress with localized, sequence-dependent structural failure to preserve B-form. *Nucleic Acids Research*, 37(16), 5568–5577. doi: [10.1093/nar/gkp556](https://doi.org/10.1093/nar/gkp556).
- Vayssières, M., Marechal, N., Yun, L., Lopez Duran, B., Murugasamy, N. K., Fogg, J. M., Zechiedrich, L., Nadal, M., & Lamour, V. (2024). Structural basis of DNA crossover capture by *Escherichia coli* DNA gyrase. *Science*, 384(6692), 227–232. doi: [10.1126/science.adl5899](https://doi.org/10.1126/science.adl5899).